SPECIES

To Cite

Jijina K, Anand PP, Neethu CB, Vardhanan YS. Geometric morphometric shape and size analysis of endemic black Clam, Villorita cyprinoides (Gray, 1825) (Mollusca: Bivalvia: Cyrenidae) from Koottayi estuary, Kerala, South India. Species 2023; 24: e33s1521

doi: https://doi.org/10.54905/disssi/v24i73/e33s1521

Author Affiliation:

Biochemistry and Toxicology Division, Department of Zoology, University of Calicut, Kerala, India

'Corresponding author

Shibu Vardhanan Y,

Biochemistry and Toxicology Division, Department of Zoology, University of Calicut, Kerala,

India

Email: svardhanan@gmail.com

ORCID List

Anand PP 0000-0002-2400-2823 Shibu Vardhanan Y 0000-0001-9820-886X

Peer-Review History

Received: 03 February 2023 Reviewed & Revised: 08/February/2023 to 06/April/2023 Accepted: 09 April 2023 Published: 14 April 2023

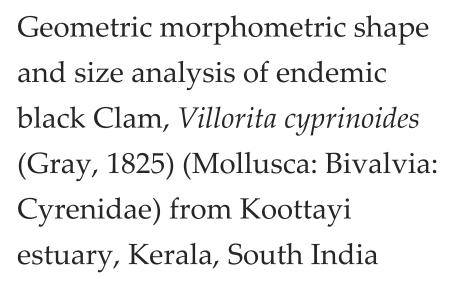
Peer-Review Model

External peer-review was done through double-blind method.

Species pISSN 2319–5746; eISSN 2319–5754



© The Author(s) 2023. Open Access. This article is licensed under a Creative Commons Attribution License 4.0 (CC BY 4.0)., which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/



Jijina K, Anand PP, Neethu CB, Shibu Vardhanan Y*

ABSTRACT

Geometric morphometric allowed us to study the shape, size and asymmetry of the right and left shells of the highly economically important endemic black clam Villorita cyprinoides (Gray, 1825) local population representing the Koottayi estuary, Tirur-Ponnani River, Kerala, South India. As far as we know, nobody has studied the population structure of black clams from southern India and this is the first geometric morphometric analysis of endemic black clam, which provided preliminary information on the population structure of black clam from the Koottayi estuary. Our study area faces a high level of pollution and anthropogenic intervention; apart from that, opens near the Arabian Sea's Barmouth. Using various multivariate statistical methods, we concluded that right and left shell halves are not of the same size and shape; and asymmetry exists. The high level of pollution and anthropogenic effect may be the reason for the existence of fluctuation asymmetry. Our research will serve as baseline information to evaluate the effects of pollution or environmental stress and human interference on endemic species. Evaluation of morphological variations works as a critical tool for future studies such as developmental instability, natural selection, phenotypic adaptation and evolution.

Keywords: Black clam, endemic species, estuary, geometric morphometrics, phenotypic plasticity, shell shape and size

1. INTRODUCTION

The fact that phenotypic variations are a reflection of adaptation or phenotypic plasticity or both. It is very challenging to explain the vital environmental selection pressure mechanisms behind phenotypic variations and how they work. Variations do not inherently inhibit adaptation due to phenotypic plasticity and promote genetic divergence in some conditions (Thimbert-Plante and Hendry, 2011; Fitzpatrick, 2012; Dowle et al., 2015). Phenotypic variations stabilize the



Species 24, e33s1521 (2023) 1 of 14

successful colonization of the individuals in a marginal environment and subsequently, contribute to genetic differentiation (Fitzpatrick, 2012). The interaction between the phenotype and the environment will contribute to the emergence of new distinct local phenotypic variations (Dowle et al., 2015; Fassatoui et al., 2019). Habitat-specific convergent evolution and phenotypic variations would be used to test adaptive hypotheses (Minards et al., 2014).

Some shelled gastropod species showed phenotypic plastic responses related to the environmental and predator pressure (Appleton and Palmer, 1988; Trussell, 2000; Doyle et al., 2010; Hollander and Butlin, 2010; Butlin et al., 2014). However, the shell characteristics can also represent a fixed genetic character (Goodfriend, 1986; Johannesson, 1996; Stankowski, 2013). Compared to linear measurements, geometric morphometrics (GMM) are now commonly used for studying the population structure of organism, particularly in inter-and intra-specific variations. Looking at the asymmetry of species is one of the fundamental aspects of GMM, which will provide valuable information on the developmental basis of morphological integration and phenotypic plasticity (Lajus et al., 2015). Mainly three different forms of asymmetry are present, such as directional asymmetry (DA), fluctuating asymmetry (FA) and antisymmetry (AS); compared to three asymmetries, FA does use as a biomarker for environmental stress assessment. Increased levels of FA seem related to various sources of stress, such as pollution (Franco et al., 2002), parasitism (Escos et al., 1995), stressful El Nino conditions (Alados et al., 1993) and poor feeding conditions (Somarakis et al., 1997). The variation in FA has already been related to a reduction in fitness performance and other population characteristics such as growth, morphology, survival, reproduction and longevity (Allenbach et al., 1999; Borrell et al., 2004; Kristoffersen and Magoulas, 2008; Arambourou et al., 2012).

Villorita cyprinoides (Gray, 1825) is an economically important endemic black clam found along the Indian coast (Jones, 1968). The black clams provide a rich and inexpensive protein source for local consumption and export purposes. Besides, shells have an immense commercial value, as they are raw materials for the manufacture of cement, calcium carbide, sand-lime bricks, distemper, glass, rayon, paper, sugar, pharmaceuticals, pesticide and poultry feed. The raw shells used in paddy fields and fish farms for neutralizing acid soil, add as slaked lime (Sudha, 1991; Joe, 1993; Kripa and Joseph, 1993; Krishna and Ammini, 2017). Clam habitat also offers many valuable ecological services; clam deposits avoid large-scale soil erosion and stabilize nearby islands (Kripa and Joseph, 1993; Krishna and Ammini, 2017). The population structure of V. cyprinoides is well known from the Vembanad lake, Kerala, however, the basic population structure of black clam from the other areas of southern India is lacking (Suja and Mohammed, 2012; Krishna and Ammini, 2017). Vembanad Lake is a transitional ecotone between the sea and the inland lake ecosystem protected by a mud bund (largest in India). It effectively prevents saline intrusion from a highly productive environment that supports the feeding, spawning and rearing of a large number of commercial fish and shellfish (ATREE, 2007). The lake bed is unsuitable for the growth of V. cyprinoides due to industrial dredging for sub-fossil lime shells and the unregulated closure of the Thanneermukkom barrage (Pillai, 1991; Laxmilatha and Appukuttan, 2002). Closing the Thanneermukkom barrage prevents the salinity intrusion into the upstream lake area that was once the highest black clam density. And it also prevents the movement of clam larvae from downstream to the upstream region of the lake. Unregulated salinity gradients and the accumulation of toxic compounds in the lake's upstream area (stagnant water type – prevent the flushing out of accumulated substances) alter the black clam habitat (Pillai, 1991; Laxmilatha and Appukuttan, 2002; Krishna and Ammini, 2017). In 2015, black clam accounted for around 75% of India's total clam production and the clam harvest decreased by 29.72% (CMFRI, 2015).

As a preliminary investigation, we selected one of Kerala's important estuarine areas, Southern India, i.e., Koottayi estuary, Tirur-Ponnani River, which is subjected to ecological deterioration due to over increasing human interventions. Koottayi estuary is opposite to Ponnani estuary and tides from the Arabian Sea influence both the estuarine system and the water is brackish almost throughout the year. Tirur-Ponnani River is joined with the Bharathapuzha River, which flows into the Arabian Sea. The river course is noted for mangroves and is home to many species of fish and migratory and native birds (DTPC, 2020). Environmental stresses, such as a change in pH, salinity, pollution, anthropogenic impacts, etc., can cause a developmental deformity in the organism and lead to a high population asymmetry. Therefore, we hypothesized that studying the endemic species' population structure from this estuarine would explain environmental health conditions' preliminary characteristics. When higher levels of asymmetry were observed in the population, it insinuated that the presence of developmental noise did trigger by environmental stress. Asymmetry analysis is a morphological marker for environmental stress assessment (Scalici et al., 2017).

This study examined the morphological shape and size of the endemic black clam, *V. cyprinoides* right and left shells from Koottayi estuary, Tirur-Ponnani River, Kerala, south India. In shell asymmetry analysis, we distinguished the right and left shells of endemic black clam and the right-left shell asymmetry. Variations in shell shape are visualized with the aid of the principal component analysis (PCA). We also recorded the size and shape covariation between the right and left shells (using 2B-PLS) and the shells' developmental covariation (PLS1 analysis).

Species 24, e33s1521 (2023) 2 of 14

2. MATERIALS AND METHODS

Sample collection

The mature organism *Villorita cyprinoides* (Gray, 1825) was collected from the Koottayi estuary (10.83'87° N, 75.55'13° E), Tirur-Ponnani River, located in the Malappuram district of Kerala, India (Figure 1A, B, C). All samples were collected during the same period (March to May, summer season, 2017); the collection was carried out with clam fishermen's help. The principal collection method is by diving underwater and collecting the clams by hand – collection estuary areas from 10 to 30 m in length and about 10 to 20 m in width and with a maximum water depth is 2.1 – 2.7 m. Specimens were transported alive to the laboratory and mechanically removed the flesh from the shell. The undamaged and clean shells were sorted and used for geometric morphometric analysis.

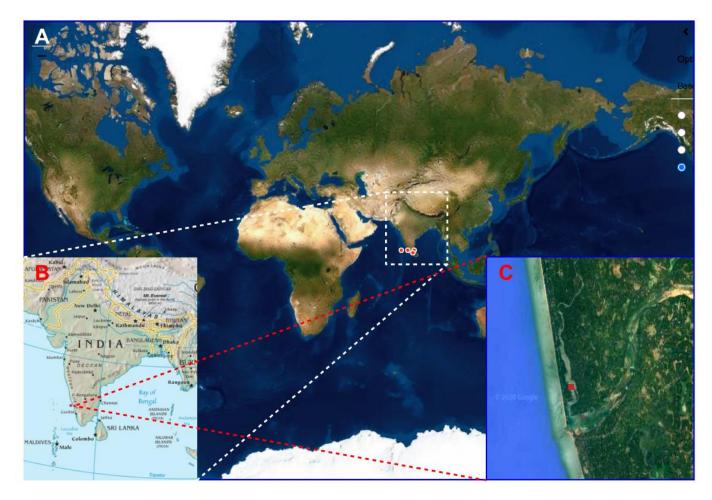


Figure 1 (A) Distribution map of *Villorita cyprinoides* – Data generated in OBIS server; (B) South India collection locality; (C) Collection site, Koottayi estuary

Geometric morphometric analysis

The collected shells were well cleaned and separated as left and right halves. A total of 101 right and 108 left ventral shell halves of *V. cyprinoides* were photographed using Canon EOS 7D (F-stop 22-27, Focal length – 180mm). Based on taxonomic characterization (Souji, 2018), 16 homologous landmarks were identified in the central region of both the left and right halves of the shell (Figure 2A, B, C) (Table 1). For land marking, 2D images of both halves of the shell were converted into tps file format using tps Utility V1.68 and tpsDig2 V2.26 used for land marking of the specimens (Rohlf, 2015). The x and y coordinates of these landmarks on the images were used to analyze asymmetry, allometry and covariations.

Species 24, e33s1521 (2023) 3 of 14

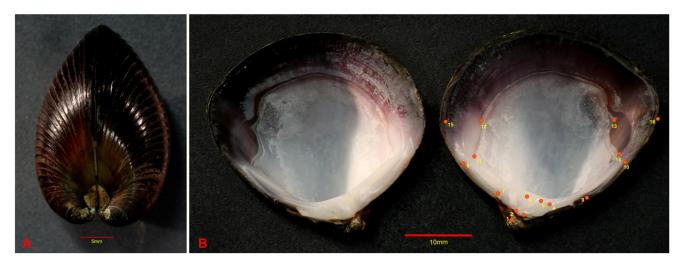


Figure 2 (A) Endemic black clam, *Villorita cyprinoides*; (B) Ventral region of both right and left halves of endemic black clam with selected geometric morphometric homologous landmarks used for shape and size analysis

Table 1 Geometric morphometric homologous landmarks used for shape and size analysis of V. cyprinoides

Landmark	Description
1	Umbo
2	Dorsal tip of Middle Cardinal Tooth
3	Dorsal tip of Posterior Cardinal Tooth
4	Antero-Ventral tip of Posterior Cardinal Tooth
5	Postero-Ventral tip of Middle Cardinal Tooth
6	Antero-Ventral tip of Middle Cardinal Tooth
7	Anterior end of Anterior Lateral Tooth
8	Posterior end of Anterior Lateral Tooth
9	Anterior end of Posterior Lateral Tooth
10	Posterior end of Posterior Lateral Tooth
11	Dorsal tip of Anterior Adductor Muscle
12	Ventral tip of Anterior Adductor Muscle
13	Ventral tip of Posterior Adductor Muscle
14	Dorsal tip of Posterior Adductor Muscle
15	Opposite end of Umbo
16	Opposite end of Ventral tip of Anterior Adductor Muscle
17	Opposite end of Ventral tip of Posterior Adductor Muscle

Statistical analysis of shape and size

In the bivalves, the right and left halves are pairs of separated structures (Savriama and Klingenberg, 2011); the analysis of fluctuation asymmetry (FA) and right-left asymmetry was based on the matching symmetry of the test organism. Therefore, to study left-right asymmetry, the reflection was removed by transforming all configurations from one body side to their mirror images (Klingenberg and Mc-Intyre, 1998). A total of three independent group analyses were performed to describe the size and shape variations, 1) the right halves population; 2) the left halves population; 3) the right-left asymmetry analysis. Following a generalized procrustes analysis (GPA), a procrustes analysis of variance (ANOVA) was used to estimate the size and shape characteristics of three groups (right, left halves and right-left). Principal component analysis (PCA) was used to detect populations' shape variations (Rohlf and Slice, 1990; Jolliffe, 2002; Uba et al., 2019).

Multivariate regression analysis was used to analyze the size-related shape changes (allometry) of right and left shell halves. In this case, the size was fixed as an independent variable and the shape as a dependent variable and using 10000 permutation analyses for testing the significance level (Monteiro, 1999; Klingenberg, 2016; Zikic et al., 2017). Discriminant function analysis (DFA) was used to visualize the asymmetry of right and left halves (group fixed as *a priori*) (Villemant et al., 2007). Size and shape

Species 24, e33s1521 (2023) 4 of 14

RESEARCH ARTICLE | OPEN ACCESS

similarities and dissimilarities between right and left halves were validated using 2B-PLS (two block-partial least square) analysis, with block one represented as size, and block two as shape (Rohlf and Corti, 2000). For developmental covariation analysis, we divided the shell into two blocks (dorsal and ventral regions), block 1 covers the umbo and cardinal tooth regions (Landmarks in block 2: 1, 2, 3, 4, 5, 6, 7, 9) and block 2 covered the shell shape regions (inner side of shell regions) (Landmarks in block 2: 8, 10, 11, 12, 13, 14, 15, 16). The covariation degree between two blocks was estimated using PLS1 within a configuration analysis with 10000 permutational analyses. The RV value below 0.5 is indicated as the covariation between the two blocks is low and above 0.5 is indicated as vice versa (Klingenberg, 2009). The entire statistical analysis was performed in MorphoJ v. 1.07a.

3. RESULTS

Size and shape variations within the population: Asymmetry

Within the right and left halves of the V. cyprinoides population do not exhibit significant shape and size variations. While comparing the left-right halves, it was proved that right and left shell asymmetry existed in the V. cyprinoides. Marginally insignificant (p = 0.052) size variation was observed in the asymmetry analysis, but they clearly show significant shape (p = 0.008) differences. Fluctuation asymmetry (FA) and directional asymmetry (DA) are only present in the right-left shape asymmetry analysis (Table 2).

Table 2 Population structure of *V. cyprinoides* from Koottayi estuary. Significant (p<0.05) result in side considered as the directional asymmetry (DA) whereas Side * Ind considered as the fluctuating asymmetry (FA)

Shell side	Analysis	Effect	SS	MS	df	F	P
Right	Centroid size	Individual	1187.941333	11.879413	100	1.00	0.4995
	Shape	Individual	0.23950893	0.000855389	2800	0.38	0.5405
Left	Centroid size	Individual	1126.308144	10.526244	107	0.91	0.8549
	Shape	Individual	0.28777594	0.0000960534	2996	0.47	0.5842
Right * Left	Centroid size	Individual	1143.315967	12.034905	95	1.05	0.0525
		Side	23.742919	23.742919	1	2.47	0.1197
		Side * Ind	914.818252	9.629666	95	1.16	0.2742
	Shape	Individual	0.237333044	0.0000892220	2660	1.11	0.0086
		Side	0.17205201	0.0061447145	28	70.15	< 0.0001
		Side * Ind	0.23300357	0.0000875953	2660	1.09	0.0288

In the discriminant function analysis (DFA), we again proved that the right and left halves are entirely separated units and do not exhibit identical shape and size patterns (Figure 3A). Using multivariate regression analysis, we demonstrated that, significant (p<0.001) size-related shape changes (allometry) have existed in both the right and left halves, but they only cover a small percentage of allometric residues. The right halves covered the 4.7945% allometric residue and the left halves covered the 3.2131% of allometric residues. In terms of size and shape similarity between the right and left halve (2B-PLS analysis) of *V. cyprinoides*, marginally significant size overlap (Block 1) was observed between the right and left halves (Figure 3B) and opposite findings were observed in the shape (Block 2) differences between the right and left halves (Figure 3C). From the overall results, we confirmed that *V. cyprinoides* right and left halves are not identical and showed significant (p<0.0001) size and shape differences.

Species 24, e33s1521 (2023) 5 of 14

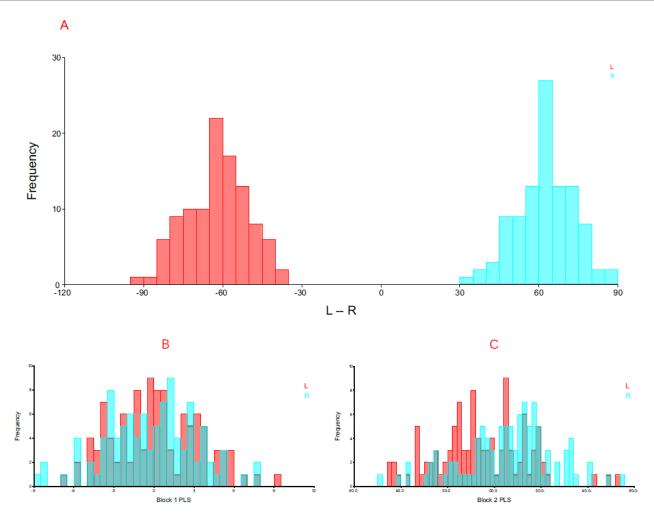


Figure 3 (A) Discriminant function analysis of right and left halves of endemic black clam; (B) Block 1 PLS size similarity and dissimilarity between right and left halves of endemic black clam; (C) Block 2 PLS shape similarity and dissimilarity between right and left halves of endemic black clam

Morphological variations of right and left shell halves

Principal component analysis (PCA) was used for quantifying the shape and size variations within the populations. We separately documented the morphological variations of the right and left halves. In the right halves, among the 28 PCs (Figure 4 A1), the first three PCs covered the 55.935% of variance (PC1 = 23.848%, PC2 = 19.766 & PC3 = 12.321%). In PC1 (Figure 4 A2), except for landmarks 6-11 & 14, all other landmarks displayed a high degree of deviation from the average point. The deviation of landmarks 12 and 15 and also 13 and 16 leads to the expansion of anterior and posterior adductor muscle scar areas. Same as in PC1 (Figure 4 A3), PC2 all landmarks showed deviation from the average position, the hinge region and umbo region landmarks pointed inward movements and also the outward movement of landmarks from the shell shape region alters the size and shape of the shell. In PC3 (Figure 4 A4), hinge region landmarks are highly conserved, they do not show substantial deviations from the average position. But the landmarks no. 10 and 8 showed too many deviations compared to other landmarks. It is indicated as the expansion of the shell in the dorsal region.

Among the 28 PCs in the left halves, the first three PCs covered 53.756% of the variance (Figure 4 B1). PC1 (Figure 4 B2) accounts the 27.210% of the variance, except for landmarks no. 8, 9 and 11, all other landmarks exhibit too many deviations from the mean shape. The same as in the right halves of PC1, the landmarks 12 and 15 (anterior adductor muscle) and 13 and 16 (posterior adductor muscle) showed too many deviations. The umbo region and cardinal tooth region landmarks moved into the inner side of shell regions. The counteract results were observed in PC2 (Figure 4 B3) and it covered the 15.850% of the variance, all landmarks were unevenly distributed in all directions, except for landmarks 7 and 10 and all other landmarks showed too many deviations from the mean position. In PC3 (10.696% of variance) (Figure 4 B4), except for umbo and cardinal tooth regions', all other regions landmarks are unevenly distributed in different directions.

Species 24, e33s1521 (2023) 6 of 14

From the overall individual population analysis (right and left halves), in the asymmetry analysis of right-left shell halves, each halve is distributed in a unique morphospace without any overlap. In the PCs morphospace distribution, the first two PCs covered 47.508% of population variance, PC1 accounted for 29.811% variance and PC2 is 17.697% of the variance (Figure 5).

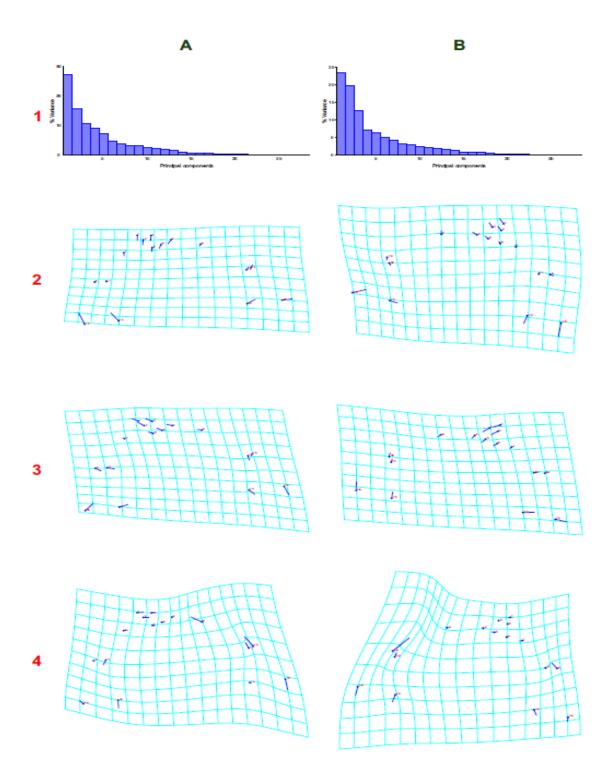


Figure 4 Principal component analysis of right and left shell halves of endemic black clam. (A1) % of variance of right shell; (B1) % of variance of left shell; A2-A4 - PC1, PC2 & PC3 of right shell; B2-B4 - PC, PC2 & PC3 of left shell

Species 24, e33s1521 (2023) 7 of 14

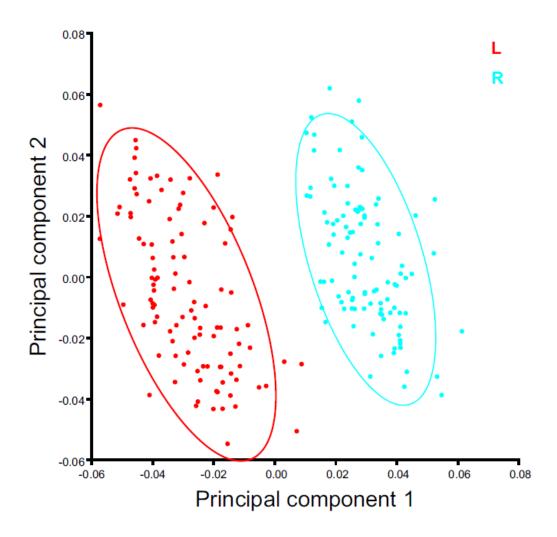


Figure 5 Morphospace shape and size distribution of right and left shell of endemic black clam

Developmental covariation analysis of V. cyprinoides

In both the right and left halves, the RV value is above 0.5; it indicated that covariation between blocks is strong. In the right halves (RV = 0.536, p<0.001) (Figure 6 A1, A2), block 1 landmarks showed a high degree of integration with the dorsal tip of the anterior adductor muscle (landmark no.11), the posterior end of the anterior lateral tooth (landmark no. 8), the ventral tip of anterior adductor muscle (landmark no.12) and the opposite end of the ventral tip of anterior adductor muscle (landmark no. 15). The landmark no. 10, 13, 14 and 16 showed less integration with other regions. In the left halves (RV = 0.567, p<0.001) (Figure 6 B1, B2), the same as in the right halves, a landmark within the block 1 displayed a high degree of covariation compared to other landmarks.

Species 24, e33s1521 (2023) 8 of 14

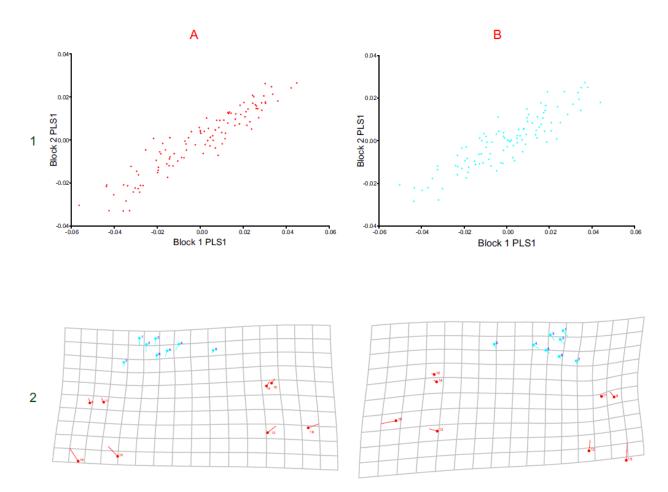


Figure 6 Developmental covariation between dorsal and ventral region of endemic black clam. (A1) % of covariation of right shell; (B1) % of covariation of left shell; (A2) Covariation between dorsal and ventral region of right shell; (B2) Covariation between dorsal and ventral region of right shell

4. DISCUSSION

Each species has its unique phenotypic variations, often related to its habitat (natural conditions – biotic and abiotic factors). During natural selection pressure, the species is forcefully to produce distinct forms (shape and size – morphological diversity) directly associated with their success and survival. Studying the morphological, phenotypic variations of local forms, would help to understand the origin of a new species (Hendry et al., 2007; Rasanen and Hendry, 2008; Dowle et al., 2015; Anand and Vardhanan, 2020). So, studying the local population phenotypic variations reveals the adaptation and stress response of an organism. Here, we describe the morphological variations of right and left shell halves of endemic black clam, *V. cyprinoides* population from Koottayi estuary, Kerala, south India.

Shell morphology is regulated by a set of heritable characters that can be shaped by environmental factors. Using GMM, we can study the shell variations into two separate components: Shape and size (Quenu et al., 2020). The shell's right and left halves did not show a significant difference in shape and size within the population. However, when considering the right and left shell asymmetry, marginally insignificant size and significant shape differences were observed. The existence of a significant shape-related FA and DA is the critical factor behind the right-left shell asymmetry. This asymmetry was suggested as the developmental instability of the local population (Koottayi estuary) of *V. cyprinoides*. Because, the FA is one of the most widely used tools for assessing developmental instability, both genotype and climatic factors regulate the developmental flux of the organisms, as evidenced by different genotypes exhibiting different levels of stability under the similar environmental conditions; and identical genotypes showing different levels of stability under varying environments (Waddington, 1942; Zakharov, 1989). Moreover, DA is developmentally regulated and likely correlated with adaptive significance, whereas FA is not associated with adaptation (Palmer, 1994). A possible implication of higher FA and developmental instability is that it can cause divergent phenotypes to express cryptic diversity (Gilbert, 2006). These phenotypes (expression of cryptic characters) are not the inheritance of acquired characters, but

Species 24, e33s1521 (2023) 9 of 14

rather a representation of an individual's existing genetic diversity that was previously impacted by the course of the developmental stability process (Nishizaki et al., 2015; Karthika et al., 2021; Anand et al., 2022). According to Woods, (2014), interactions between environmental factors and developmental noise can contribute to a higher range of phenotypes. This diversity arises from the stochastic processes of cell division and development, not from plasticity or adaptation. The fundamental mechanisms that induce the minor changes in FA-related morphology remain unclear. The stochastic switching of gene expression is the driving mechanism behind an organism's developmental instability (Klingenberg, 2003).

The black clam attains sexual maturity at a length of 11 to 15 mm (0.4 to 0.6 in). It does not show sex reversal or hermaphroditism. It is spawned twice a year, from May to August and from January to late March. The most important factor that causes spawning is the change in salinity, though the temperature is not a factor. The optimum spawning salinity is around 10-12 ppt (Arun, 2009; Ravindran et al., 2006). According to the National Green Tribunal (NGT), Kerala pollution control board, Govt. of India, Southern Zone (2016), reported that the Tirur-Ponnani River and Koottayi estuary were confronted with very serious environmental culmination, such as dumping of garbage, solid waste materials, oil spills, biomedical waste, etc. Apart from this, the Koottayi estuary is near the Ponnani harbor; the high level of anthropogenic influences has also changed the natural condition of the Koottayi estuary. Irregular water current from the Arabian Sea, high levels of pollution and human interference alter the native biodiversity of the Koottayi estuary (Sreenisha and Paul, 2016). These factors change the salinity, temperature and biodiversity of the estuary; it will ultimately lead to the developmental noise of organisms living in the estuary. We concluded that the high level of pollution and human intrusions is the crucial reason behind the FA of the black clam population of the Koottayi estuary. There is little information available on the functional aspects of FA in bivalves. Asymmetry in bivalve shells is known to cause drag (Olivera and Wood, 1997) and many thus increase susceptibility to wave action. Shell asymmetry can also increase desiccation, which is known to increase with valve gaps (Kennedy, 1976). Mismatches in valve size can make mussels more vulnerable to decapod predators crushing techniques (Elner, 1978). Further research is currently required to determine the functional implications of shell asymmetry.

From the multivariate regression analysis, significant allometry has existed in both the right and left halves of *V. cyprinoides*, even though it covers a low percentage of allometric residues. Our findings corroborate the previous results of Suja and Mohamed, (2012), according to this author, the biometric documentation of *V. cyprinoides* perfectly expressed the length-width and length-height relationships (positive allometric) even though they are collected from different stations of Vembanad Lake, Kerala, India. The finding implies that the growth in width and height is superior to an increase in length. This kind of positive allometry has been observed in some other bivalves, such as *Perna viridis* (Narasimham, 1981), *Donax Semistriatus*, *Spisula solida*, *S. subtruncata* and *Ensis siliqua* (Gaspar et al., 2002). The shell morphology variation analysis based on PCA revealed that each PC of the right and left shell exhibits different grades of variations, suggesting that a morphologically clear-cut differentiation existed in both the right and left halves of black clam. And these morphometric shape variations could be used for distinguishing the right and left halves of a black clam. In general, changes in the relative proportion of bivalve shells during growth are related to maintaining a suitable physiological ratio concerning environmental conditions (Rhoads and Pannella, 1970). Bivalve shells become progressively higher and wider during ontogeny to counter involuntary dislodgement, turbulence and currents (Hinch and Bailey, 1988). Shell morphology and the selective proportions of many bivalves are known to affect several environmental factors, such as latitude (Beukema and Meeham, 1985), depth and type of bottom (Claxon et al., 1998), tidal level (Dame, 1972), sediment type (Newell and Hidu, 1982) and burrowing behaviour (Seed, 1980).

Between primary and higher trophic levels species, benthic organisms constitute a critically important food chain. They change the bottom ecosystem structurally and maintain the biogeochemical cycling of nutrients and other substances. Because of their restricted mobility, benthic invertebrates are vulnerable to pollution and other environmental disturbances. In corollary, they were used as indicators for ecological health in general and focus on a wide range of environmental impact assessments (Sushama, 2014). Unraveling shell shape and size variations in a local environment reflect the natural selection pressure at work rather than random genetic drift (Giokas et al., 2014). From the 2B-PLS covariation study, we can confirm that the shape variable is more affected than size. And marginally significant covariation (dorsal and ventral region of the shell) was observed only in the PLS1 analysis. The covariation analysis (2B-PLS and PLS1) proved that shape-related developmental instability existed in *V. cyprinoides*. We previously stated that the Koottayi estuary is confronted with a high level of pollution and human interferences, altering the embryonic and post-embryonic development of black clam.

Species 24, e33s1521 (2023) 10 of 14

5. CONCLUSION

From the overall findings, two key conclusions can be drawn to explain the shell shape and size variations of *V. cyprinoides* from the Koottayi estuary: (1) this phenotypic variation can be the starting point for the expression of the cryptic diversity of black clam associated with their natural selection pressure; and (2) shell usually function as a highly integrated unit, reduction of covariation between the anterior and posterior shell regions; and high level of alteration of size and shape may lead to the extinction species from the Koottayi estuary. More genomic, reproductive, developmental and population studies are required to address our work's main findings. As a future recommendation, more sampling from other local populations across southern India is required to establish the evolution of natural selection and also to help to classify the native population. The analysis of an endemic organism's size, shape and asymmetry can effectively assess environmental health status. Our research will serve as a framework for future studies to determine the effect of pollution or environmental stress and human intrusion on endemic species and morphological variations can be used as a key tool for future studies.

Acknowledgements

We thank the Department of Zoology, University of Calicut for providing the infrastructural facility. The present study was financially supported by UGC-SAP (F.3-6/2012 (SAP-II) dated 10.10.2012).

Disclosure statement

No potential conflict of interest was reported by the authors.

Informed consent

Not applicable.

Ethical approval

The ethical guidelines are followed in the study for species observation & identification.

Conflicts of interests

The authors declare that there are no conflicts of interests.

Funding

The study has received external funding from UGC-SAP (F.3-6/2012 (SAP-II) dated 10.10.2012).

Data and materials availability

All data associated with this study are present in the paper.

REFERENCES AND NOTES

- Alados CL, Escos J, Emlen JM. Developmental instability as an indicator of environmental stress in the Pacific hake *Merluccius productus*. Fish Bull 1993; 91:587-593.
- Allenbach DM, Sullivan KB, Ludy MJ. Higher fluctuating asymmetry as a measure of susceptibility to pesticides in fishes. Environ Toxicol Chem 1999; 18:899-905.
- Anand PP, Seena S, Peter J, Vardhanan YS. Detection of geographical specific plasticity and the effect of natural selection pressure on the wing size and shape of *Bactrocera dorsalis* (Diptera: Tephritidae). Biologia 2022; 77(4). doi: 10.10 07/s11756-022-01059-x
- Anand PP, Vardhanan YS. Computational modelling of wet adhesive mussel foot proteins (Bivalvia):

- Insights into the evolutionary convolution in diverse perspectives. Sci Rep 2020; 10:2612. doi: 10.1038/s41598-020-59169-y
- Appleton RD, Palmer AD. Water-borne stimuli released by predatory crabs and damaged prey induce more predator-resistant shells in a marine gastropod. Proc Natl Acad Sci U S A 1988; 85:4387-4391.
- Arambourou H, Beisel JN, Branchu P, Debat V. Patterns of fluctuating asymmetry and shape variation in *Chironomus riparius* (Diptera: Chironomidae) exposed to nonylphenol or lead. PLoS One 2012; 7:e48844.
- 7. Arun AU. An assessment on the influence of salinity in the growth of black clam (*Villorita cyprinoides*) in a

Species 24, e33s1521 (2023) 11 of 14

- cage in Cochin Estuary with a special emphasis on the impact of Thanneermukkom salinity barrier. Aquac Aquar Conserv Legis Int J Bioflux Soc 2009; 2:433-447.
- ATREE. Conserving biodiversity and sustaining our future. Biennial Report 2005-2007. Bangalore, India: Ashoka Trust for Research in Ecology and the Environment 2007.
- Beukema JJ, Meehan BW. Latitudinal variation in linear growth and other shell characteristics of Macoma balthica. Mar Biol 1985; 90:27-33.
- Borrell YJ, Pineda H, Mc-Carthy I, Vazquez E, Sanchez JA, Lizana GB. Correlations between fitness and heterozygosity at allozymes and microsatellite loci in the Atlantic salmon, *Salmo salar* L. Heredity 2004; 92:585-593.
- 11. Butlin RK, Saura M, Charrier G, Jackson B, André C, Caballero A, Coyne JA, Galindo J, Grahame JW, Hollander J, Kemppainen P, Martínez-Fernández M, Panova M, Quesada H, Johannesson K, Rolán-Alvarez E. Parallel evolution of local adaptation and reproductive isolation in the face of gene flow. Evolution 2014; 68:935-949.
- Claxon WT, Wilson AB, Mackie GL, Boulding EG. A genetic and morphological comparison of shallow and deep-water populations of the introduced Dreissenid bivalve *Dreissena bugensis*. Can J Zool 1998; 76:1269-1276.
- 13. CMFRI. Annual report 2015-2015. Cochin, India: Central Marine Fisheries Research Institute 2015.
- 14. Dame RF. Comaprison of various allometric relationships in intertidal and subtidal American oysters. Fish Bull 1972; 70:11 21-1126.
- 15. Dowle EJ, Morgan-Richards M, Brescia F, Trewick SA. Correlation between shell phenotype and local environment suggests a role for natural selection in the evolution of *Placostylus* snails. Mol Ecol 2015; 24:4205-4221. doi: 10.1111/mec.13302
- 16. Doyle S, Mac-Donald B, Rochette RM. Is water temperature responsible for geographic variation in shell mass of *Littorina obtusata* (L.) snails in the Gulf of Maine? J Exp Marine Biol Ecol 2010; 394:98-104.
- DTPC. District tourism promotion council, Malappuram 2020. https://www.malappuramtourism.org/rivers-backwaters.php.
- 18. Elner RW. The mechanics of predation by the shore crab, *Carcinus maenas* (L.), on the edible mussel, *Mytilus edulis* L. Oecologica 1978; 36:333-344.

- 19. Escos J, Alados CL, Emlen JM, Alderstein S. Developmental instability in the Pacific hake parasitized by myxosporeans *Kudoa* spp. Trans Am Fish Soc 1995; 124:943-945.
- 20. Fassatoui C, Jenhani ABR, Romdhane MS. Relative growth, shell morphology and genetic relationships between freshwater mussels of the genus *Unio* (Mollusca: Bivalvia: Uninoidea) from rivers of the Ichkeul watershed (Tunisia). Molluscan Res 2019; 39(4):1-12. doi: 10.1080/13235818.2019.16 31958
- 21. Fitzpatrick BM. Underappreciated consequences of phenotypic plasticity for ecological speciation. Int J Ecol 2012; 2012:12.
- 22. Franco A, Malavasi S, Pravoni F, Nasci C, Torricelli P. Ethoxyresorufin O-deethylase (EROD) activity and fluctuating asymmetry (FA) in *Zosterisessor ophiocephalus* (Teleostei: Gobiidae) as indicators of environmental stress in the Venice Lagon. J Aquat Ecosyst Stress Recovery 2002; 9:239 -247.
- Gaspar MB, Santos MN, Vasconcelos P, Monteiro CC. Shell morphometric relationships of the most common bivalve species (Mollusc: Bivalvia) of the Algarve cost (southern Portugal). Hydrobiologia 2002; 477:73-80.
- 24. Gilbert SF. Developmental biology. Sinauer, Sunderland, Massachusetts, USA 2006.
- 25. Giokas S, Pall-Gergely B, Mettouris O. Nonrandom variation of morphological traits across environmental gradients in a land snail. Evol Ecol 2014; 28:323-340.
- Goodfriend GA. Variation in Land-snail shell form and size and its causes: A Review. Syst Biol 1986; 35:204-223.
- 27. Hendry AP, Nosil P, Rieseberg LH. The speed of ecological speciation. Funct Ecol 2007; 21:455-464.
- 28. Hinch SG, Baiiley RC. Within and among-lake variations in shell morphology of the fresh water clam *Elliptio complanta* (Bivalvia: Uninodae) from south central Ontario Lakes. Hydrobiologia 1988; 157:27-32.
- 29. Hollander J, Butlin R. The adaptive value of phenotypic plasticity in two ecotypes of a marine gastropod. BMC Evol Biol 2010; 10:333.
- 30. Joe KK. Observations on the population characteristics of the Corbiculid clam *Villorita cyprinoides* (Gray) in the chitrapuzha portion of the Vembanad Lake. MSc Thesis submitted to Cochin University of Science and Technology 1993.
- 31. Johannesson B. Population differences in behaviour and morphology in the snail *Littorina saxatilis*:

Species 24, e33s1521 (2023) 12 of 14

- Phenotypic plasticity or genetic differentiation? J Zool 1996; 240:475-493.
- 32. Jolliffe LT. Principal component analysis. 2nd edition. Springer. New York, USA 2002.
- 33. Jones S. The molluscan fishery resources of India. Mar Biol Ass India 1968; 3:906-918.
- 34. Karthika K, Anand PP, Seena S, Vardhanan YS. Wing phenotypic plasticity, quantitative genetics, modularity and phylogenetic signal analysis revealed the niche partitioning in two fruit fly species, *Bactrocera dorsalis* and *Zeugodacus cucurbitae*. Int J Trop Insec Sci 2021; 42:1487-1504. doi: 10.1007/s42690-021-00668-4
- Kennedy VS. Desiccation, higher temperatures and upper intertidal limits of three species of sea mussels (Mollusca: Bivalvia) in New Zealand. Mar Biol 1976; 35:127-137.
- 36. Klingenberg CP, Mc-Intyre GS. Geometric morphometric of developmental instability: Analyzing patterns of fluctuating asymmetry with Procrustes methods. Evolution 1998; 52(5):13 63-1375.
- 37. Klingenberg CP. A developmental perspective on developmental instability: Theory, models and mechanisms. Polak M (editor), Developmental instability: Causes and consequences. Oxford University Press, New York, USA 2003.
- Klingenberg CP. Morphometric integration and modularity in configurations of landmarks: Tools for evaluating a priori-hypotheses. Evol Dev 2009; 11:405-421.
- 39. Klingenberg CP. Size, shape and form: Concepts of allometry in geometric morphometrics. Dev Genes Evol 2016; 226(3):113-137.
- 40. Kripa V, Joseph S. Clam fishery of north Vembanad Lake. Mar Fish Infor Serv T & E Ser 1993; 119:5-11.
- 41. Krishna AV, Ammini P. Population characteristics of *Villorita cyprinoides* in Vembanad Lake, India. Lake Reserv Manag 2017; 34(2):130-140.
- 42. Kristoffersen JB, Magoulas A. Investigating anchovy (*Engraulis encrasicolus* L.) population structure in the Mediterranean Sea using multiple methods. Fish Res 2008; 91: 187-195.
- 43. Lajus D, Katolikov M, Strelkov P, Hummel H. Fluctuating and directional asymmetry of the blue mussel (*Mytilus edulis*): Improving methods of morphological analysis to explore species performance at the Northern Border of its range. Symmetry 2015; 7:488-514. doi: 10.3390/sym7020488

- 44. Laxmilatha P, Appukuttan KK. A review of the black clam (*Villorita* cyprinoides) fishery of the Vembanad Lake. Indian J Fish 2002; 49:85-91.
- 45. Minards NA, Trewick SA, Godfrey AJR, Morgan-Rchards M. Convergent local adaptation in size and growth rate but not metabolic rate in a pair of parapatric Orthoptera species. Biol J Linn Soc 2014; 113:123-135.
- 46. Monteiro IR. Multivariate regression models and geometric morphometrics: The search for casual factors in the analysis of shape. Syst Biol 1999; 48:192-199.
- 47. Narasimham KA. Dimensional relationships and growth of green mussel *Perna viridis* in Kakinada Bay. Indian J Fish 1981; 28:240-248.
- 48. National Green Tribunal, Gov. of India, Southern Zone, Chennai. Application no. 308 of 2013 (SZ) (THC) (WP No. 27528 of 2012, High court of Kerala) 2016. http://www.india environmentportal.org.in/files/Tirur%20Ponnani%20 River%20pollution%20NGT%20Order%20Kerala.pdf
- 49. Newell CR, Hidu H. The effects of sediment type o growth rate and shell allometry in the soft-shelled clam *Mya arenaria* (L.). J Exp Mar Biol Ecol 1982; 65:285-295.
- 50. Nishizaki MT, Barron S, Carew E. Thermal stress increases fluctuation asymmetry in marine mussels: environmental variation and developmental instability. Ecosphere 2015; 6(5): 85. doi: 10.1890/ES14-00399.1
- 51. Olivera A, Wood W. Hydrodynamics of bivalve shell entrainment and transport. J Sediment Res 1997; 67:514-526.
- 52. Palmer AR. Fluctuating asymmetry analysis: A primer. In Developmental Instability: Its origins and evolutionary implications. Markow TA (editor.). Springer, Dordrecht, the Netherlands 1994.
- 53. Pillai VK. Studies on the hydrobiology and pollution of the Vembanad Lake and adjacent water (Dissertation), Cochin, India: Cochin University of Science and Technology 1991.
- 54. Quenu M, Trewick SA, Brescia F, Morgan-Richards M. Geometric morphometrics and machine learning challenge currently accepted species limits of the land snail *Placostylus* (Pulmonata: Bothriembryonidae) on the Isle of Pines, New Caledonia. J Molluscan Stud 2020; 1-7. doi: 10.1093/mollus/ey z031
- 55. Rasanen K, Hendry AP. Disentangling interactions between adaptive divergence and gene flow when

Species **24**, e33s1521 (2023)

- ecology drives diversification. Ecol Lett 2008; 11:624-636.
- 56. Ravindran K, Appukuttan KK, Sivassankara-Pillai VN, Boopendranath MR. Report on the committee of experts on ecological and environmental impact of degrading at Vaduthala Kayal and Vaikkam Kayyal. Unpublished Rep. submitted to the Government of Kerala, Thiruvananthapuram 2006; 45.
- 57. Rhoads DC, Pannella G. The use of molluscan growth patterns in ecology and paleoecology. Lethaia 1970; 3:143-161.
- 58. Rohlf FJ, Corti M. The use of two-block partial squares to study covariation in shape. Syst Biol 2000; 49:740-753.
- 59. Rohlf FJ, Slice D. Extensions of Procrustes method for the optimal superimposition of landmarks. Syst Zool 1990; 39(1): 40-59.
- 60. Rohlf FJ. The tps series of software. Hystrix Italian J Mammal 2015; 26:9-12.
- 61. Savriama Y, Klingenberg CP. Beyond bilateral symmetry: Geometric morphometric methods for any type symmetry. BMC Evol Biol 2011; 11(1):1-24.
- 62. Scalici M, Traversetti L, Spani F, Malafoglia V, Colamartino M, Persichini T, Cappello S, Mancini G, Guerriero G, Colasanti M. Shell fluctuating asymmetry in the sea-dwelling benthic bivalve *Mytilus galloprovincialis* (Lamarck, 1819) as morphological markers to detect environmental chemical contamination. Ecotoxicol 2017; 26:396-404. doi: 10.1007/s1064 6-017-1772-9
- 63. Seed R. Shell growth and forms in Bivalvia. In: Skeltal growth of aquatic organisms. Biological Records of Environmental change (editors. Rhoads DC, Lutz KA) Plenum Press, New York 1980; 23-67.
- 64. Somarakis S, Kostikas I, Peristerakis N, Tsimenides N. Fluctuating asymmetry in the otoliths of larval anchovy *Engraulis encrasicolus* and the use of developmental instability as an indicator of condition in larval fish. Mar Ecol Prog Ser 1997; 151:191-203.
- 65. Somarakis S, Kostikas I, Tsimenides N. Fluctuating asymmetry in the otoliths of larval fish as an indicator condition: Conceptual and methodological aspects. J Fish Biol 1997; 51:30-38.
- Souji S. Molluscan fauna of southeast coast of India with special references to Bivalvia. PhD thesis. University of Kerala 2018.
- 67. Sreenisha KS, Paul PT. An assessment of the pollution and its impact on the diversity of phytoplankton in Tirur River, Malappuram district,

- Kerala, India. Int J Curr Mirobiol Appl Sci 2016; 5(7):180-190.
- 68. Stankowski S. Ecological speciation in an island snail; evidence for the parallel evolution of a novel ecotype and maintenance by ecologically dependent postzygotic isolation. Mol Ecol 2013; 22:2726-2741.
- 69. Sudha BN. Breeding biology of Villorita cyprinides (Gray) in relation to salinity gradients. MSc Thesis submitted to Department of fisheries, College of fisheries, Cochin, Kerala Agriculture University 1991.
- 70. Suja N, Mohamed KS. Biometric relationships of *Villorita cyprinoides* (Gray) from Vembanad Lake, Kerala, India. Asian Fish Sci 2012; 25:29-39.
- 71. Sushama S. Biodiversity of Ponnani estuary, Kerala. J Aquat Biol Fish 2014; 2:785-791.
- 72. Thimbert-Plante X, Hendry AP. The consequences of phenotypic plasticity for ecological speciation. J Evol Biol 2011; 24:326-342.
- 73. Trussell GC. Phenotypic clines, plasticity and morphological trade-offs in an intertidal snail. Evolution 2000; 54:151-166.
- 74. Uba KIN, Monteclaro HM, Noblezada-Payne MMP, Quinitio GF. Sexual dimorphism, asymmetry and allometry in the shell shape of *Modiolus metcalfei* (Hanley, 1843) collected from Dumangas, IIoilo, Philippines: A geometric morphometric approach. Comput Ecol Softw 2019; 9(3):107-120.
- 75. Villemant C, Simbolotti G, Kenis M. Discrimination of *Eubazus* (Hymenoptera, Braconidae) sibling species using geometric morphometrics analysis of wing venation. Syst Entomol 2007; 32(4):625-634. doi: 10.1111/j.1365-3113.2007.00389.x
- Waddington CH. Canalization of development and the inheritance of acquired characters. Nature 1942; 150:563-565.
- 77. Woods HA. Mosaic physiology from developmental noise: Within-organism physiological diversity as an alternative to phenotypic plasticity and phenotypic flexibility. J Exp Biol 2014; 217:35-45.
- 78. Zakharov VM. Future Prospects for population phenogenetics: Psychology and general sociology reviews. Taylor & Francis 1989.
- 79. Zikic V, Stankovic SS, Petrovic A, Milosevic M, Tomanovic Z, Klingenberg CP, Ivanovic A. Evolutionary relationship of wing venation and wing size and shape in Aphidiinae (Hymenoptera: Braconidae). Org Divers Evol 2017; 17:607-617. doi: 10.1007/s13127-017-0338-2

Species 24, e33s1521 (2023) 14 of 14